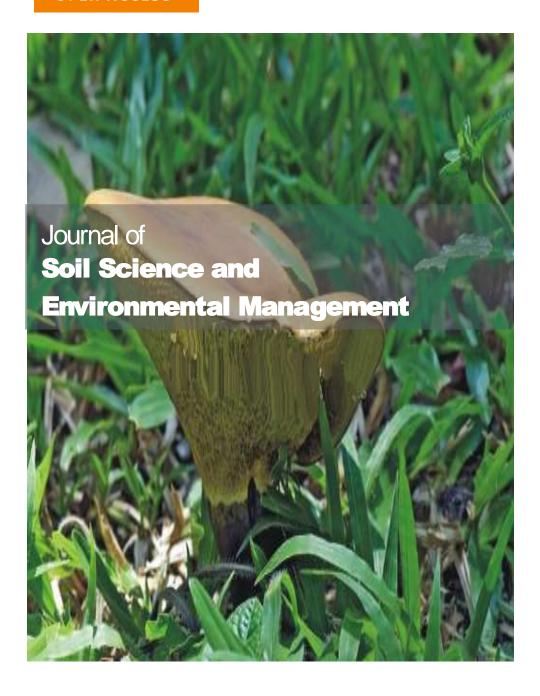
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## Journal of Soil Science and Environmental Management

#### Full Length Research Paper

# Influence of phosphorus and potassium fertilizers on growth and yield of potato (*Solanum tuberosum* L.) at Assosa, Benishangul Gumuz Regional State, Western Ethiopia

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Potato is one of the most important food security and cash crops in Ethiopia. It is constrained by poor soil fertility. A field experiment was conducted at Assosa Agricultural Research Centre to investigate the effect of phosphorus and potassium fertilizers on the growth performance and yield of potato. The experiment was laid out as a randomized complete block design (RCBD) in 4×6 factorial arrangement of potassium (0, 100, 200 and 300 kg K<sub>2</sub>O ha<sup>-1</sup>) and phosphorus (0, 46, 92, 138, 184 and 230 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup>) in three replicates. A potato variety, Gudanie (CIP-386423-13) was used. Analysis of the data revealed that the interaction effect of both phosphorus and potassium did not influence the phonotypic, growth parameters and tuber yields of potato, but their main effect they significantly influenced days to 50% flowering, physiological maturity, plant height , marketable and total tuber yields, leaf area, above and underground dry biomasses. Optimum above and underground dry biomass (232.11 and 494.74 Mg\* ha<sup>-1</sup>), marketable (23.94 kg K<sub>2</sub>O ha<sup>-1</sup>) and total tuber (29.56 kg K<sub>2</sub>O ha<sup>-1</sup>) yields were attained at 200 kg K<sub>2</sub>O ha<sup>-1</sup>; for phosphorus, optimum marketable tuber (23.30 Mg ha<sup>-1</sup>), total tuber (28.83 Mg ha<sup>-1</sup>), and yield of above ground and underground dry matter (218.48 and 479.60 Mg ha<sup>-1</sup>) were attained at 138 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup>. The lowest yield obtained from above ground and underground dry matter, marketable and total tuber in both fertilizers were recorded at zero level.

**Key words:** Fertilizer rate, phenotypic parameter, plant height, biomass, tuber yield, 1 Mg (Megagram) = 1000 kg.

#### INTRODUCTION

Potato (*Solanum tuberosum* L.) is an important food and cash crop in Eastern and Central Ethiopia. The food potential of the potato crop has been indicated in literature as being a cheap source of human diet. Potato produces 74.5% more food energy per unit area than

wheat and 58% more than rice. Also it produces 54% more protein per acre than wheat and 77% more than rice (Thornton and Sieczka, 1980). It has relatively high carbohydrate and quality protein, and low fat content (FAO, 1980; Dean, 1994). Due to its shallow root system

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and short crop duration, the nutrient requirement of potato for growth and development is very high, especially due to its coarse roots and sparse root hairs, making potato inefficient in the uptake of phosphorus (Perrenoud, 1993; Nigussie-Dechassa et al., 2003). Unfortunately, P is one of the least accessible nutrients in most soil especially under tropical conditions where low P availability is a big challenge to agricultural production (Kochian *et al.*, 2004). Phosphorus is present to some extent in all soils and unlike nitrogen it is held tightly by soil particles and therefore it is not easily leached from the soil (Carroll and Reiley, 2011).

On the other hand, uptake of fertilizer nutrients (NPK) by potato per unit area and time is quite high due to fast rate of early growth and tuber bulking (Singh, 1999). Potato is less efficient user of potassium than other crops (Trehan and Claassen, 2000). A potato crop of average yield in the tropics may remove 50-80 kg N ha<sup>-1</sup>, 20-30 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> and 80-100 kg K<sub>2</sub>O ha<sup>-1</sup> from the soil (Sikka, 1982). The fertilizer use efficiency of K in potato ranges between 50 and 60% (Bansal and Trehan, 2011).

Many researchers reported that phosphorus application affects crop growth by increasing radiation interception (over the whole season) or by increasing light use efficiency (Plenet et al., 2000). Plants grown under low P level develop lower plant biomass due to either limited light interception or the amount of absorbed photosynthetically active radiation (PAR) (Colomb et al., 1995) to a less efficient conversion of the intercepted radiation (Plenet et al., 2000). The reduced total leaf area could be due to both reduced number of leaves and smaller individual leaf size. A decrease in number of leaves in P deficient plants can be ascribed to reduced leaf initiation and activity of the shoot meristems (Chiera et al., 2002). On the other hand, the reduced individual leaf size can be due to reduced cell division rate (Assuero et al., 2004) or reduced epidermal cell expansion (Radin and Eidenbock, 1984), which ultimately affect leaf expansion rate. Colomb et al. (2000) observed significantly lower final leave number in non-P treated plants than in P treated ones, which ultimately affected total plant leaf area. Likewise, many researchers reported that potassium application caused a significant increase in vegetative growth (Li-Xiu and Liu-Ya, 2003; Pauletti and Menarim, 2004). The vegetative growth gradually and significantly increased by increasing level of potassium application (Asmaa and Hafez, 2010).

Moreover, fertilizer requirement varies across locations due to many reasons such as difference in soil types, nutrient availability of the soil, economic factors of the area, moisture supply, and variety (Getu, 1998; IAR, 2000). The P adsorption and fixation is influenced by soil pH, clay type and content, as well as the amount of iron and aluminium oxides (Mesifin, 1998). There is a claim that potassium is least deficient in Ethiopian soils and that the soils have good potassium supplying power (Tsedale, 1983, Tekalign and Haque, 1988). However,

this claim may not apply to acidic soils (Brich, 1969; Atanasiu, 1970). The highly weathered soils have low exchangeable potassium (below 0.3 milliequivalents per 100 g of soil) (Murphy, 1963; Mesfin, 1998). Results from analysis of soils of Assosa Research Agricultural Center (AsARC) indicated very low available exchangeable potassium (0.85)ppm and 0.157 milliequivalents per 100 g, respectively). These values are very low according to Landon (1991) and Mengel and Kirkby (2001). In general, the soils in the south-western and western Ethiopia are acidic due to the high precipitation that leads to lose of basic cations by leaching (Mesfin, 1998). Thus, contrary to the general belief that most soils of Ethiopia are rich in potassium, this nutrient is likely limited in acidic soils due to high rate of leaching (Mengel and Kirkby, 2001).

Hence, in view of the fact that Ethiopian soils are poor in fertility and have problem of low soil pH, phosphorus fixation, and N and K leaching, and realizing the importance of fertilizers in potato production, the use of inorganic and organic fertilizers in potato production for optimum yield and quality tuber production is vital. This research is, therefore, aimed at investigating the effect of different rates of inorganic phosphorus and potassium fertilizers on the phenology, growth and yield of potato under Assosa condition.

#### **MATERIALS AND METHODS**

#### **Experimental site**

The experiment was carried out in the Benishangul Gumuz Region of Ethiopia at Assosa Agricultural Research Center, which is located at latitude of 10°02' N, longitude of 34°34' E and an altitude of about 1553 m above sea level. The area has a mean annual rainfall of 1100 mm. It has a warm humid climate with mean maximum and minimum annual temperatures of 32 and 17°C, respectively (AsARC, 2011). The soil of the area is Nitosol, which is characteristically reddish to brown in colour. It is acidic having a pH of 5.1 and silty in texture with contents of 49% silt, 17% sand, and 34% clay. The soil has organic matter content of 4.86%, and total nitrogen, available phosphorus and exchangeable potassium contents of 0.068%, 8.52 mg kg¹ soil and 0.136 cmol kg¹ soils, respectively, at 0-30 cm soil depth.

#### **Experimental materials**

#### Planting material

The potato variety called 'Gudanie' (CIP-386423-13) was used as a planting material which has wide-range environmental adaptation in Ethiopia. It requires up to 120 days for physiological maturity and considered moderately resistant to the late blight disease (Woldegiorgis et al., 2008).

#### Fertilizer material

Triple superphosphate (TSP) (46%  $P_2O_5$ ) and potassium chloride (KCI) (60%  $K_2O$ ) were used as sources of phosphorus and potash, respectively. Urea (CO[NH<sub>2</sub>]<sub>2</sub>) (46% N) was used as a source of

nitrogen.

#### Treatments and experimental design

The treatments consisted of four levels of potassium (0, 100, 200 and 300 kg K<sub>2</sub>O ha<sup>-1</sup>), and six levels of phosphorus (0, 46, 92, 138, 184 and 230 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup>). The basis for these levels was the pretesting of the soil nutrient which was low in available phosphorus (8.52 ppm) and very low in exchangeable potassium (0.12 cmol kg soil) according to Mengel and Kirkby (2001). The experiment was laid out as a randomized complete block design (RCBD) in a 4 x 6 factorial arrangement and replicated three times. There were 24 treatment combinations, which were assigned to each plot randomly. The total number of plots was 72 and each plot had a gross area of 11.25 m<sup>2</sup> with 3 m length and 3.75 m width. Each plot contained five rows of potato plants, with each row accommodating 10 plants per row with a total population of 50 plants per plot at the spacing of 0.75 m and 0.30 m between rows and plants, respectively. The spacing between plots and adjacent blocks was 1 m and 2 m, respectively.

#### **Experimental procedures**

#### Land preparation

The land was prepared in May to June 2011 using a tractor and human labour. Ridges on which to plant the tubers were constructed manually.

#### **Planting**

Medium-sized (40 to 60 g) and sufficiently sprouted potato tubers (with 2 to 3 cm long sprouts) were planted on ridges at the specified spacing on 08 July, 2011.

#### Fertilizer application

Application of phosphorus and potash fertilizers at the specified rates was done by banding the granules of the two fertilizers at the depth of 5 to 10 cm below and around the seed tuber at planting. All phosphorus was applied at planting while potash was applied in two splits [1/2 at emergence and 1/2 at mid-stage of the plant (at about 40 days after planting)] because of the problem of leaching caused by high rainfall. Nitrogen at the blanket recommended rate of 92 kg N ha<sup>-1</sup> was applied to all plots equally in the form of urea in three splits [1/4<sup>th</sup> at planting, 1/2 at mid-stage of the plant (at about 40 days after planting), and 1/4<sup>th</sup> at the initiation of tubers (at the start of flowering)].

#### Other cultural practices

Weeds were controlled by hoeing. Earthing-up was done as required to prevent exposure of tubers to direct sunlight, to promote tuber bulking and to ease harvesting. Mancozeb ( $C_8H_{12}MnN_4S_8Zn$ ), active ingredient of maneb and metiram (USEPA, 2005), was sprayed at the rate of 50 g per 20 L of water to control late blight disease.

#### **Data collection**

Data were recorded on different phenotypic and growth characteristics as well as yield of potato.

#### Crop phenotype

Days to 50% flowering was referred to the time required to attain 50% of the plant to flower while days to 50% physiological maturity was referred to the time required by the plant to reach the stage of growth when 50% of the vines started senescing. This was done when haulms (vines) of 50% of the plant population became yellow or the leaves senesced according to IBPGR descriptor list (IBPRG, 1977).

#### Plant growth parameters

#### Plant height (cm)

Plant height (cm) is the height from the base to the apex of the plant. It was determined by measuring the height of 20 randomly selected plants using a ruler from the central three rows at flowering.

#### Leaf area index (LAI)

Leaf area index (LAI) was obtained by dividing the value of the leaf area by the area of the land occupied by the plant using the following formula (Diwaker and Oswalt, 1992):

$$Leaf area index (LAI) = \frac{LA_m \times N}{A}$$

Where;  $LA_m$  = mean leaf area of the plant (cm²); A = the area (cm²) occupied by one plant in the cropping area; and N = number of leaves on the plant. To determine the total leaf area (cm²), five plants (hills) from plot was randomly selected, tagged, and the leaf length (LL) (cm) of the individual plants at 20 days intervals measured. Individual leaf area (LA) of the potato plants (cm²) was estimated from individual leaf length using the following formula developed by Firman and Allen (1989):

$$\log_{10}(LA) = 2.06 \times \log_{10}(LL) - 0.458$$

#### Total dry biomass (Mg ha<sup>-1</sup>)

Total dry biomass (Mg ha<sup>-1</sup>) was referred to the dry weight of leaves, stems, roots, stolons, and tubers. It was determined from 10 randomly taken plants from the central rows just before senescence (at physiological maturity). Samples of dry weights were taken after air-drying and oven-drying the samples at 65°C till constant weight is obtained (CIP, 1984).

#### Tuber dry matter content (%)

Five fresh tubers were randomly selected from each plot and weighed. The tubers were then sliced and dried in an oven at 65°C until a constant weight was obtained and the dry weight was recorded. The dry matter percent was calculated according to the following formula (Williams and Woodbury, 1968):

Dry matter (%) = 
$$\frac{\text{Wieght after drying (g)}}{\text{Initial weight (g)}} \times 100$$

#### Yield parameters

Marketable tuber yield (kg ha<sup>-1</sup>) was the weight of tubers which are free from diseases, insect pests, and greater than or equal to 25 g

**Table 1.** Mean squares of potato phenology, growth parameters and yield components as influenced by phosphorus, potassium and their interaction.

Variable		Р	K	РхК
Degree of freedom (d.f)		5	3	15
Dhanalagical narameter	Days to 50% flowering	4.6**	71.2**	1.0 <sup>Ns</sup>
Phenological parameter	Days to 50% physiological maturity	28.5**	566.9**	3.4 <sup>Ns</sup>
	Plant height (cm)	386.10**	941.83**	74.66 <sup>Ns</sup>
	Leaf area index	2.51Ns	25.90**	2.87 <sup>Ns</sup>
Growth parameter	Above ground dry biomass (Mg ha <sup>-1</sup> )	723.65*	26843.41**	1179.08 <sup>Ns</sup>
	Underground dry biomass (Mg ha <sup>-1</sup> )	21415.76*	108768.76**	11313.30 <sup>Ns</sup>
	Total dry biomass (Mg ha <sup>-1</sup> )	27381.16*	243102.38**	17602.55 <sup>Ns</sup>
Tubanidalda	Marketable (Mg ha <sup>-1</sup> )	53.89**	166.37**	15.09 <sup>Ns</sup>
Tuber yields	Total (Mg ha <sup>-1</sup> )	60.94**	230.19**	19.36 <sup>Ns</sup>

<sup>\*\*, \*:</sup> Significant differences at 1 and 5% level of significance, respectively; Ns = non-significant at 5% level of significance; P = phosphorus  $(P_2O_5)$ ; K = potassium  $(K_2O)$ .

in weight was recorded. Unmarketable tuber yield (kg ha<sup>-1</sup>) includes the weight of tubers that are diseased and/or rotten and small-sized (less than 25 g in weight) were recorded. Total tuber yield (kg ha<sup>-1</sup>) is the sum of tuber yield weights of marketable and unmarketable tubers.

#### Statistical analysis

Data were subjected to analysis of variance (ANOVA) according to the Generalized Linear Model (GLM) of SAS version 9.0 (SAS Institute, 2004). Significant differences between treatment means were separated using the least significance difference (LSD) test at 5% significance level.

#### **RESULTS**

#### Influence of P and K on Phenological parameters

The analysis of variance for P and K interactive effect was non-significant while the main effect of phosphorus and potassium fertilizers showed highly significant difference (P < 0.01) on days to flowering and maturity (Tables 1 and 2). Increasing phosphorus application from nil to 92 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> prolonged the days required by the potato plants to attain 50% flowering by about 2.7%. Increasing the phosphate supply further to 184 and 230 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> further prolonged the days required by the potato plants to attain 50% flowering to about 2.9 and 3.3%, respectively, as compared to plants grown in the control treatment (Table 2). However, plants grown at the phosphate supply of 92 and 138 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> had values of days to 50% flowering that were in statistical parity (Table 2). Similarly, application of phosphorus fertilizer prolonged the time required by the potato crop to reach physiological maturity (Table 2). Thus, compared to plants that received no phosphorus fertilizer, plants that received phosphorus at the maximum rate of 230 kg ha<sup>-1</sup> required about three more days (about 3%) to attain 50% physiological maturity. Even if 230 kg ha<sup>-1</sup> delayed the 50% flowering to 3 days, the treatments that received beyond 46 kg ha<sup>-1</sup> did not show statistical significant.

Application of potassium fertilizer linearly and highly significantly prolonged the days required to reach 50% flowering and physiological maturity. The days to 50% flowering was delayed by about 9% when the rate of potassium was increased from nil to 300 kg  $\rm K_2O~ha^{-1}$ . Similarly, the days for 50% physiological maturity were delayed by about 13.89% (about 14 days) in response to increasing the rate of potassium from nil to 300 kg  $\rm K_2O~ha^{-1}$  (Table 2).

#### Influence of P and K on growth parameters

#### Plant height

Phosphorus and potassium did not interact to influence plant height; however, it responded highly significantly (P < 0.01) to the main effects of phosphorus and potassium application rates (Tables 1 and 3). Plants grown in the control treatment were highly significantly shorter than plants grown at the rates of  $\geq 92 \text{ kg P}_2\text{O}_5 \text{ ha}^{-1}$ . Increasing the rate of phosphorus from nil to to 92, 138, 184 and 230 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> resulted in highly significant increases in plant height (Table 3) However, the heights of plants grown in plots supplied with only 46 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> were in statistical parity with the heights of plants in the control treatment. Similarly, the plant height of potato was highly affected by the main effect of potassium application (Table 1); however, the application of potassium fertilizer above the rate 100 kg K<sub>2</sub>O ha<sup>-1</sup> did not respond statistically to plant height parameter (Table 3).

Table 2. Phenological parameters of potato as affected by phosphorus and potassium fertilizers application.

Treatment	Days to 50% flowering	Days to 50% physiological maturity
P₂O₅ (kg ha <sup>-1</sup> )		
0	52.3°	99.0 <sup>e</sup>
46	53.1 <sup>bc</sup>	99.8 <sup>de</sup>
92	53.7 <sup>ab</sup>	100.6 <sup>cd</sup>
138	53.7 <sup>ab</sup>	101.3 <sup>bc</sup>
184	54.0 <sup>a</sup>	102.0 <sup>ab</sup>
230	53.8 <sup>ab</sup>	103.3 <sup>a</sup>
F-test	**	**
LSD (5%)	0.9	1.3
K₂O (kg ha <sup>-1</sup> )		
0	51.2 <sup>d</sup>	94.3 <sup>d</sup>
100	52.6 <sup>c</sup>	99.1°
200	54.3 <sup>b</sup>	103.2 <sup>b</sup>
300	55.7 <sup>a</sup>	107.4 <sup>a</sup>
F-test	**	**
LSD (5%)	0.7	1.03
CV (%)	2.69	1.52

Means followed by the same letter within a column are not significantly different at 5% level of significance; \*\* = significant at P < 0.01 probability level; LSD = Least significant difference; CV = Coefficient of variation.

Table 3. Growth parameters of potato as influenced by phosphorus and potassium application at Assosa.

Treatment	Plant height	Leaf area	Aboveground dry biomass	Underground dry biomass	Total dry biomass
	(cm)	index	(Mg ha <sup>-1</sup> )	(Mg ha <sup>-1</sup> )	(Mg ha <sup>-1</sup> )
P₂O₅ (kg ha <sup>-1</sup> )					
0	56.56 <sup>d</sup>	3.80	162.77 <sup>b</sup>	346.41 <sup>b</sup>	509.18 <sup>b</sup>
46	61.04 <sup>cd</sup>	3.75	169.03 <sup>b</sup>	431.84 <sup>ab</sup>	600.87 <sup>ab</sup>
92	67.14 <sup>ab</sup>	3.67	177.18 <sup>ab</sup>	425.92 <sup>ab</sup>	603.09 <sup>ab</sup>
138	71.91 <sup>a</sup>	3.75	218.48 <sup>a</sup>	479.60 <sup>a</sup>	698.08 <sup>a</sup>
184	65.87 <sup>bc</sup>	3.92	212.90 <sup>a</sup>	477.18 <sup>a</sup>	690.08 <sup>a</sup>
230	69.66 <sup>ab</sup>	3.83	211.50 <sup>a</sup>	485.32 <sup>a</sup>	696.82 <sup>a</sup>
F-test	**	Ns	*	*	*
LSD (5%)	5.95	0.61	41.495	84.466	115
K₂O (kg ha <sup>-1</sup> )					
0	54.65 <sup>b</sup>	4.77 <sup>c</sup>	89.34 <sup>c</sup>	330.06 <sup>c</sup>	419.39 <sup>c</sup>
100	67.44 <sup>a</sup>	6.41 <sup>b</sup>	186.71 <sup>b</sup>	423.46 <sup>b</sup>	610.17 <sup>b</sup>
200	69.12 <sup>a</sup>	7.09 <sup>ab</sup>	232.11 <sup>a</sup>	494.74 <sup>a</sup>	726.86 <sup>a</sup>
300	70.25 <sup>a</sup>	7.48 <sup>a</sup>	259.75 <sup>a</sup>	515.91 <sup>a</sup>	775.66 <sup>a</sup>
F-test	**	**	**	**	**
LSD (5%)	4.87	0.84	33.88	68.97	93.90
CV (%)	11.07	19.44	22.72	25.34	22.93

Means of the same main effect followed by the same letter or with no superscript letter within a column are not significantly different at 5% level of significance; \*\* = significant at P < 0.01 probability level; \* = significant at P < 0.05 probability level; Ns = non-significant at P < 0.05 probability level; LSD = Least significant difference; and CV = Coefficient of variation.

#### Leaf area index (LAI)

The analysis of variance of the influence of phosphorus

and potassium on leaf area index (LAI) is shown in Table 1. Application of potassium highly significantly (P < 0.01) influenced LAI of potato while phosphorus fertilization

and its interaction with potassium did not affect this parameter. Increasing potassium fertilization from nil to 100 kg increased leaf area index of the crop by about 34%. Further increased to 100 and 200 kg  $K_2O$  ha<sup>-1</sup> increased the leaf area by about 49 and 57% as compared to that of the control. However, the leaf area index recorded at 100 and 200 kg  $K_2O$  ha<sup>-1</sup> as well as that recorded at 200 and 300 kg  $K_2O$  ha<sup>-1</sup> were in statistical parity (Table 3).

#### Dry biomass

The two nutrients did not interact to influence the dry aboveground biomass, underground as well as total biological dry mass of potato. However, the main effect of both phosphorus and potassium significantly (P < 0.05) and highly significantly (P < 0.01) affected all the aforementioned three parameters, respectively (Tables 1 and 3). Increasing the rate of phosphorus from 0 to 46 or 92 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> did not significantly change the aboveground dry biomass yield. However, when the rate of phosphate was further increased to 138 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup>, the above ground dry biomass yield increased by about 34% but it did not respond to phosphorus fertilizer application beyond this level (Table 3). In case of underground dry biomass yield of potato, inconsistence increment of yield of dry biomass appeared as the application of phosphorus increases. All levels of phosphorus except the control had no statistically significance response on these parameters (Table 3). While in case of the influence of potassium, increasing the rate of potassium resulted in significantly increased aboveground. underground as well as total dry biomass yield of the crop even more vigorously than the increases recorded in response to phosphorus application. All treatments that received potassium fertilizer gave a significant better above-and underground biomass as well as total biomass compared to the control treatment; however, there was no significant biomass yield in treatments that received 200 and 300 kg K<sub>2</sub>O ha<sup>-1</sup>.

## Influence of P and K on marketable and total tuber yields

The interaction of the phosphorus and potassium nutrients did not influence both marketable and total tuber yields. However, phosphorus highly significantly affected marketable and total tuber yields (Tables 1 and 4). Both marketable and total tuber yields obtained in the control treatment were highly significantly lower than those received rates of  $\geq$  46 kg  $P_2O_5$  ha $^{-1}$ , but the tuber yields did not respond statistically up to the level of 184 kg  $P_2O_5$  ha $^{-1}$ . Similarly, the main effect of potash significantly influenced marketable, as well as total tuber yields of potato (Tables 1 and 4). The lowest marketable and total

tuber yields were also obtained from the control treatment of potassium. However, the application of potassium rate 100 and 200 kg  $K_2O$  ha<sup>-1</sup> did not respond statistically to marketable tuber yield. Likewise, application beyond 200 kg  $K_2O$  ha<sup>-1</sup> did not increase marketable and total tuber yields statistically (Table 4).

#### DISCUSSION

#### Influence of P and K on phenological parameters

Plants that received phosphorus prolonged the 50% flowering and physiological maturity of about 3%, as compared to plants that did not receive phosphorus fertilizer (Table 4). The longer duration required for flowering and maturity in response to the increased rates of phosphorus application could be ascribed to beneficial effect of phosphate fertilizer on growth which could be explained in terms of enhanced early canopy growth and increased radiation interception for photosynthesis (Jenkins and Ali, 1999). On the other hand, it might be the synergetic effect of phosphorus and potassium with the nitrogen uptake which enhanced the vegetative stage and hence, delayed flowering and maturity since its uptake was enhanced as the uptake of these nutrients increased. The result of this study is consistent with that of Zelalem et al. (2009) who observed that phosphorus fertilization significantly prolonged days required for flowering and to attain physiological maturity in potato. The observations of the current investigation, however, are in contrast to those of Kleinkopf et al. (1987) and Armstrong (1999) where phosphorus nutrient was reported to be associated with shortening maturity.

Application of potassium fertilizer linearly and highly significantly prolonged the days required to reach 50% flowering and physiological maturity. The longer duration required for maturity in response to the increased rates of potassium application could be ascribed to favourable growth conditions and less interplant competition at higher levels of potassium which may have prolonged the developmental stage for higher starch accumulation and partitioning to the tubers. This result coincides with that of Harris (1978) who noted that potassium application prolonged the leaf area duration and, thus the days required to reach physiological maturity.

#### Influence of P and K on growth parameters

#### Plant height

The result showed that potato plants grown at the rates of  $\geq 92 \text{ kg P}_2\text{O}_5 \text{ ha}^{-1}$  had statistically longer height than the control. Increasing the rate of phosphorus from nil to 92, 138, 184, and 230 kg  $\text{P}_2\text{O}_5$  ha $^{-1}$  resulted in highly significant increases in plant height by about 19, 27, 16,

**Table 4.** Tuber yield parameters and harvest index of potato as influenced by phosphorus and potassium application at Assosa during the main cropping season in 2011.

Treatment	Marketable tuber yield(Mg ha <sup>-1</sup> )	Total tuber yield ( Mg ha		
P₂O₅ (kg ha <sup>-1</sup> )	·			
0	18.83 <sup>c</sup>	23.32 <sup>c</sup>		
46	22.51 <sup>b</sup>	26.63 <sup>b</sup>		
92	21.70 <sup>b</sup>	26.31 <sup>b</sup>		
138	23.30 <sup>ab</sup>	28.83 <sup>ab</sup>		
184	23.00 <sup>ab</sup>	27.16 <sup>ab</sup>		
230	25.24 <sup>a</sup>	29.80 <sup>a</sup>		
F-test	**	**		
LSD (5%)	2.36	2.65		
K₂O (kg ha <sup>-1</sup> )				
0	18.17 <sup>c</sup>	22.06 <sup>c</sup>		
100	22.47 <sup>b</sup>	26.71 <sup>b</sup>		
200	23.94 <sup>ab</sup>	29.56 <sup>a</sup>		
300	25.14 <sup>a</sup>	29.70 <sup>a</sup>		
F-test	**	**		
LSD (5%)	1.93	2.17		
CV (%)	12.82	11.95		

Means of the same main effect followed by the same letter within a column are not significantly different at 5% level of significance, DMRT test; \*\* = significant at P < 0.01 probability level; \* = significant at P < 0.05 probability level; Ns = non-significant at P < 0.05 probability level; LSD = Least significant difference; CV = Coefficient of variation.

and 23%, respectively (Table 3). Similarly, the heights of plants grown in plots supplied with only 46 kg  $P_2O_5$  ha<sup>-1</sup> were in statistical parity with the heights of plants in the control treatment. This might indicate that phosphorus was still sub-optimal for growth of the plants to full height at this rate. However, the rates of phosphorus over 92 kg  $P_2O_5$  ha<sup>-1</sup> showed statistically parity in response to plant height. This indicates that the rate of phosphorus for growth of the potato plants to optimum height was 92 kg  $P_2O_5$  ha<sup>-1</sup>. The present finding agrees with that of Grewal et al. (1991) who reported that potato plant heights were positively related to phosphorus fertilizer applications in phosphorus deficient soils.

application of potassium Similarly, significantly enhanced the height of potato plant. When the rate of potash was increased from nil to 100 kg K<sub>2</sub>O ha<sup>-1</sup>, the plant height was increased by about 23%. However beyond the rate of 100 kg K<sub>2</sub>O ha<sup>-1</sup>, plant height was nonsignificantly affected (Table 3). This shows that potassium also contributes to increased cell division and elongation whereby it results in higher canopy development. This suggestion is in line with that of Marschner (1995) who reported that potassium results in enhanced cellular growth and development. The result of the present investigation is consistent with the findings of Asmaa and Hafez (2010) who noted that application of higher rates of potassium resulted in higher plant height of potato. In addition, Khandakhar et al. (2004) reported that application of potassium significantly increased plant height.

#### Leaf area index

The higher leaf area index obtained in response to increased potassium application could be attributed to enhanced growth of vegetative plant parts due to the simulative effect of the increased supply of the nutrient on assimilate synthesis and meristematic growth of tissues, which may have resulted in more number of leaves and higher leaf area indices. The leaf area index value obtained in this study is inconsistent with the suggestion of Marschner (1995) who stated that leaf area index of potato for optimum yield ranges between three and six. The results obtained from this study are in accord with those of Al-Moshileh et al. (2005) who reported that potassium is important for plant growth partly due to its effect on LAI and consequently light interception and dry matter production. The higher nutrient uptake right from early stage of crop growth was one of the reasons for improved vegetative growth at higher levels of potassium supply. However, some reports revealed that leaf area index was affected also by phosphorus (Yong-fu et al., 2006).

#### Dry biomass

Increasing the rate of phosphate from 0 to 46, and 92 kg

P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> did not affect total dry biomass yield of the crop. However, increasing the rate of the nutrient from nil to 138 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> increased this parameter by about 37% (Table 3). Total dry biomass yield did not increase significantly beyond this level of the supply of the nutrient. The increase in dry matter production of the plant in response to phosphorus application could be attributed to increased radiation interception (over the whole season) or increased light use efficiency, and hence, increased canopy growth and increased conductance of the plant that enhanced photoassimilation and production of dry matter. The results obtained in this experiment are in accord with that of Ali and Anjum (2004) who reported that increased phosphorus supply increased dry matter production in plants. Similarly, Soltanpour and Cole (1978) found that application of phosphorus fertilizers increased leaf, stem and tuber growth rates and, consequently dry matter and yields. Consistent with the results of this study. Ali and Anium (2004) reported that higher rates of phosphorus application resulted in increased total dry weight and there was some evidence from ground cover scores that leaf senescence occurred earlier at higher phosphorus rates. Similarly, Zelalem et al. (2009) also reported that above and underground biomass yields increased significantly in response to the application of phosphorus fertilization. This may be attributed to enhanced interception of radiation and enhanced leaf expansion and photosynthesis especially during the early phase of

Increasing the rate of potassium resulted in significantly increased aboveground, underground as well as total dry biomass yield of the crop even more vigorously than the increases recorded in response to phosphorus application. As increasing the rate of potash from 0 to 100 kg K<sub>2</sub>O ha<sup>-1</sup>, already significantly increased aboveground, underground, and total dry biomass yields by about 109, 28, and 45%, in the order cited. Similarly, increasing the rate of the nutrient from 100 to 200 kg K<sub>2</sub>O ha<sup>-1</sup> significantly further increased the aboveground, underground, and total dry biomass yields of the crop by about 24, 17, and 19%, respectively. Beyond this level of potash supply, no significant increases in all three parameters were recorded (Table 3). The increase in biological dry masses in response to the increased levels of potassium might be attributed to the fact that the nutrient enhanced growth of more vegetative parts including plant height, branches, total leaf area and production of more tubers through promoting enzymatic activities and enhancing the translocation of assimilates and protein synthesis as described by Devlin and Witham (1986). These results obtained in this study are in accord with those reported by Asmaa and Hafez (2010) who noted that a higher application of potassium resulted in higher biomass production in potato. Potassium increases leaf expansion particularly at early stages of growth, extends leaf area duration by delaying leaf

shedding near maturity. It increases both the rate and duration of tuber bulking. Its application activates a carbohydrate metabolism and proteins and assists in the translocation of carbohydrates from leaves to tubers (Imas and Bansal, 1999).

## Influence of P and K on marketable and total tuber yields

Increasing phosphorus application from nil to 46 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> significantly increased marketable tuber yield by about 20%. Increasing the rate of phosphorus further from 46 to 92, 138, and 184 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> did not affect marketable tuber yield. However, when the rate of phosphate was increased from 0 to 230 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup>, marketable tuber yield increased by about 34%. Similarly, increasing the rate of phosphorus from nil to 46 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> significantly increased total tuber yield by about 14%. Increasing the rate of the nutrient from 46 to 92, 138, and 184 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> did not affect total tuber yield. However, when the rate was increased from 0 to 230 kg P<sub>2</sub>O<sub>5</sub> ha<sup>1</sup>, total tuber yield increased by about 28%. In general, the result revealed that optimum marketable as well as total tuber yields were attained at 138 kg  $P_2O_5$  ha<sup>-1</sup> (Table 4). The marketable and total tuber yields increased in response to the application of phosphorus fertilizer possibly due to increased radiation interception and increased conversion efficiency. Corroborating this result, Zameer et al. (2010) showed significant increases in potato tuber yields in response to increased phosphorus application due to increased radiation interception rather than increased conversion efficiency. This is also in line with what Allison et al. (2001) suggested that the increased ground cover and radiation interception observed was the mechanism through which phosphorus fertilizer increased potato tuber yields.

Similarly, increasing the rate of potassium from 0 to 100 kg K<sub>2</sub>O ha<sup>-1</sup> increased marketable and total tuber yields by about 24 and 21%, respectively. Besides, further increasing the rate of the nutrient from 100 to 200 K<sub>2</sub>O ha<sup>-1</sup> increased both tuber yields by about 7 and 11%, respectively. Increasing the rate of the mineral fertilizer from 200 to 300 kg K<sub>2</sub>O ha<sup>-1</sup> did not change marketable and total tuber yields. In general, the amount of mineral potassium fertilizer that optimized marketable and total tuber yields amounted to 200 kg K<sub>2</sub>O ha<sup>-1</sup> (Table 4). The results of this study are corroborated by those of Al-Moshileh et al. (2005) who reported that marketable tuber yield increased significantly in response to increased potassium application rates. Besides, Khandakhar et al. (2004) and Asmaa and Hafez (2010) reported significant increments in yield due to potassium application. However, Mulubrhan (2004) and Zelalem et al. (2009) found that there was no significant increment in marketable as well as total tuber yields of potato in response to increasing the rate of potassium.

#### Conclusion

phosphorus and potassium affected phonological parameters (days to flowering and days to maturity). The application of 184 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup>, and 230 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup> prolonged days to 50% flowering (by about 3%), and physiological maturity (by about 4%), respectively, as compared to the control while application of potassium at 300 kg K<sub>2</sub>O ha<sup>-1</sup> delayed the days required to reach 50% flowering by about 9% and physiological maturity by 13% (about 13 days) as compared to the days required by plants grown in the control treatment to reach the same stage of growth. However, phosphorus and potassium did not interact to influence these phenological parameters.

All growth parameters, plant height, leaf area index and biological dry mass were highly significantly affected by the main effect of potassium. The optimum plant height (67.44 cm) was recorded at the level of 100 kg K<sub>2</sub>O ha<sup>-1</sup>, leaf area (7.09), and above ground (232.12 Mg ha<sup>-1</sup>), underground (494.74 Mg ha<sup>-1</sup>) and total (726.86 Mg ha<sup>-1</sup>) dry biomasses at the level of 200 kg K<sub>2</sub>O ha<sup>-1</sup>. Similarly, the main effect of phosphorus significantly influenced the plant height and the mentioned dry biomasses of potato. However, the main effect of phosphorus did not affect leaf area. Generally, the optimum plant height (67.14 cm), and above (218.48 Mg ha<sup>-1</sup>), underground (479.60 Mg ha<sup>-1</sup>) and total (698.08 Mg ha<sup>-1</sup>) dry biomasses were recorded at the level of 138 kg P<sub>2</sub>O<sub>5</sub> ha<sup>-1</sup>. Moreover, phosphorus and potassium did not interact to affect these growth parameters.

Application of both of these fertilizers also highly significantly influenced the marketable and total tuber yields and their application beyond 138 kg  $P_2O_5$  ha<sup>-1</sup> and 200 kg  $K_2O$  ha<sup>-1</sup> had non-significant increment on tuber yield. Generally, in response to phosphorus, the optimum marketable (23.30 Mg ha<sup>-1</sup>) and total tuber (28.83 Mg ha<sup>-1</sup>) yields were attained at the level of 138 kg  $P_2O_5$  ha<sup>-1</sup> while in case of potassium the optimum marketable (23.94 Mg ha<sup>-1</sup>) and total tuber (29.56 Mg ha<sup>-1</sup>) yields were attained at the application of 200 kg  $K_2O$  ha<sup>-1</sup>.

#### CONFLICT OF INTERESTS

The authors have not declared any conflict of interests.

#### **REFERENCES**

- Ali H, Anjum MA (2004). Aerial growth and dry matter production of potato (Solanum tuberosum L.) cv. Desiree in relation to phosphorus application. International Agricultural Biological Journal 6(3):458-461.
- Allison JH, Flower JH, Allen EJ (2001). Effects of soil and foliar-applied phosphorus fertilizers on the potato (*Solanum tuberosum* L.) crop. Agricultion Science Journal 137:379-395.
- Al-Moshileh AM, Errebhi MA, Motawei MI (2005). Effect of various potassium and nitrogen rates and splitting methods on potato under sandy soil and arid environmental conditions. Emirate Agricultural Science Journal 17(1):1-9.

- Armstrong DL (1999). Effect of phosphorus on crop maturity. Better Crops 83:14-19.
- Assosa Agriculture Research Center (AsARC) (2011). Assosa Agricultural Research Center metrological data for 2011. Assosa, Ethiopia.
- Asmaa, R.M. and M.M. Hafez, 2010. Increasing productivity of potato plants (*Solanum tuberosum* L) by using potassium fertilizer and humic acid application. Int. Academic Research Journal 2(2):83-88.
- Assuero SG, Mollier A, Pellerin S (2004). The decrease in growth of phosphorus deficient maize leaves is related to a lower cell production. Plant Cell and Environment 27:887-895.
- Atanasiu N (1970). Response to K by cereals on different Ethiopian Soils. In: Proc. of the 9<sup>th</sup> Congress of the Inter. Potash Institute of Potassium Sym. on the Role of Fertilization in the Intensification of Agricultural Production. International Potato Institute, Berne, Switzerland pp. 249-257.
- Bansal SK, Trehan SP (2011). Effect of potassium on yield and processing quality attributes of potato. *Karnataka* Agricultural Science Journal 24(1):48-54.
- Brich HF (1969). Agricultural chemistry and soil science. pp. 81-96. In: Progress Report for the Period March 1968 to 1969. Holleta Genet Research Station, IAR, Addis Ababa, Ethiopia.
- Carroll LS, Reiley HE (2011). Introductory Horticulture, 8<sup>th</sup> Ed. Delmar, Cengage Learning, Clifton Park, New York pp. 41-59.
- Chiera J, Thomas J, Rufty T (2002). Leaf initiation and development in soybean under phosphorus stress. Experimental Botany Journal 53:473-481.
- Colomb B, Bouniols A and Delpech C (1995). Effect of various phosphorus availabilities on radiation use efficiency in sunflower biomass until anthesis. Plant Nutrition Journal 18:1649-1658.
- Colomb B, Kiniry JR, Debaeke P (2000). Effect of soil phosphorus on leaf development and senescence dynamics in field-grown maize. Agronomy Journal 92:428-435.
- Dean BB (1994). Managing the Potato Production System. Food Products Press, USA 61p.
- Devlin RM, Witham FH (1986). Plant Physiology, 4<sup>th</sup> Edition. CBS Publ. and Distr., Delhi, India.
- Diwaker B, Oswalt DL (1992). Research planning and data handling. ICRISAT, Andra Pradesh, India. 89p.
- Food and Agriculture Organization (FAO) (1980). Production year book. Rome, Italy.
- Firman DM, Allen EJ (1989). Estimating individual leaf area of potato from leaf length. Agricultural Science Journal, 112:425-426.
- Getu B (1998). Yield, quality, and nitrogen uptake of potato (*Solanum tuberosum* L.) as influenced by rate and time of nitrogen application. An M.Sc. Thesis Presented to the School of Graduate Studies of Alemaya University. Ethiopia 101p.
- Grewal JS, Trehan SP, Sharma RC (1991). Phosphorus and potassium nutrition of potato. CPRI Technical Bulletin No.31.Central Potato Research Institute, Shimla, HP, India 43p.
- Harris PM (1978). Mineral Nutrition. pp. 195-243. In: P.M. Harris, (ed.). The potato Crop: The Scientific Basis for Improvement. Chapman and Hall. London.
- Institute of Agricultural Research (IAR) (2000). Holetta Guenet Research Station Progress Report. April 2000. EIAR, Addis Ababa, Ethiopia.
- International Potato Centre (CIP) (1984). Potato for the developing world. Lima, Peru 150p.
- International Board for Plant Genetic Resources (IBPGR) (1977). Descriptors for the Cultivated Potato. IBPGR, Rome, Italy.
- Imas P, Bansal SK (1999). Integrated nutrition management in potato.
   In: Proc. Sym. of Global Potato Meet, Central Plant Research Institute, December, New Delhi.
- Jenkins PD, Ali H (1999). Growth of potato cultivars in response to application of phosphate fertilizer. Annual Applied Biology 135: 431-38.
- Khandakhar SM, Rahman MM, Uddin MJ, Khan SA, Qudduus KG (2004). Effect of lime and potassium on potato in acid soils. Pakistan Biology Science Journal 7(3):380-383.
- Kleinkopf GE, Westermann DT, Willie MJ (1987). Specific Gravity of Russet Burbank Potatoes. American Potato Journal 64:579-587.
- Kochian LV, Hoekenga OA and Pineros MA (2004). How do crop plants

- tolerate acid soils? Mechanisms of aluminum tolerance and phosphorus efficiency. Annual. Review of Plant Biology 55:459-493.
- Landon JR (1991). Booker Tropical Soil Manual: Handbook for Soil Survey and Agricultural Land Evaluation in the Tropics and Sub Tropics. Longman Inc. New York.
- Li-Xiu M, Liu-Ya P (2003). An experiment on the best application amount of K<sub>2</sub>SO<sub>4</sub> for potato (*Solanum tuberosum* L.) grown in Chernozem soil. Chinese Potato Journal 17(1):23-24.
- Marschner H (1995). Functions of mineral nutrients: macronutrients, pp. 299-312. In: H. Marschner (ed.). Mineral Nutrition of Higher Plants, 2<sup>nd</sup> Edition. Academic Press New York.
- Mengel K, Kirkby EA (2001). Principles of Plant Nutrition, 5<sup>th</sup> Edition. Dordrecht: Kluwer Academic Publishers 849p.
- Mesfin A (1998). Nature and Management of Ethiopian Soils. Alemaya University of Agriculture, Ethiopia pp. 227-246.
- Mulubrhan H (2004). The effects of nitrogen, phosphorus, and potassium fertilization on the yield and yield components of potato (*Solanum tuberosum* L.) grown on vertisols of Mekelle area, Ethiopia. An M.Sc Thesis Presented to School of Graduate Study of Haramaya University, Ethiopia.
- Murphy HF (1963). Fertility and Other Data on Some Ethiopian Soils. College of Agriculture and Mechanical Art, Alemaya, Ethiopia 48 p.
- Nigussie-Dechassa N, Schenk MK, Steingrobe N (2003). Phosphorus efficiency of cabbage (*Brassica oleraceae* L. var. Capitata), carrot (*Daucus carota* L.), and potato (*Solanum tuberosum* L). Plant and Soil 250:215-224.
- Pauletti V, Menarim E (2004). Timing, sources and rates of potassium fertilizer application for potato. *Scientia Agraria* 5(1/2):15-20.
- Perrenoud S (1993). Fertilizing for High Yield Potato, 2<sup>nd</sup> Edition. Inter. Potash Institute, Basel, Switzerland.
- Plenet D, Mollier A, Pellerin S (2000). Growth analysis of maize field crops under phosphorus deficiency, II. Radiation-use efficiency, biomass accumulation and yield components. Plant and Soil 224:259-272
- Radin JW, Eidenbock MP (1984). Hydraulic conductance as a factor limiting leaf expansion of phosphorus-deficient cotton plants. Plant Physiology 75:372-377.
- Statistical Analysis System Institute (SAS) (2004). SAS statistical guide for personal computers, version 9.0. SAS Institute.
- Sikka L (1982). Fertilizer and manure requirement of the potato, In: Nganga S andShideler F (eds), Potato Seed Production for Tropical Africa. International Potato Center, Lima, Peru pp. 92-95.
- Singh JP (1999). Potassium fertilization of potatoes in north India. In: Proc. of IPI Workshop on Essential Role of K in Diverse Cropping Systems, held at the 16<sup>th</sup> World Congress of Soil Science, Montpellier, France, 20-26 August 1998. Inter. Potash Institute, Basel, Switzerland pp. 123-127.
- Soltanpour PN, Cole CV (1978). Ionic balance and growth of potatoes as affected by N plus P fertilization. *American* Potato Journal 55:549-60.
- Tekalign M, Haque I (1988). Potassium status of some Ethiopian soils. East African Agri. and Forestry Journal 53:123-130.
- Thornton RE, Sieczka JB (1980). Commercial potato production in North America. American Potato Journal 57:534-536.
- Trehan SP, Claassen N (2000). Potassium uptake efficiency of potato and wheat in relation to growth in flowing solution culture. Potato Research Journal 43:9-18.
- Tsedale W (1983). Potassium nutrition of potato (*Solanum tuberosum* L.) on four major Hararge highland soils. M.Sc. Thesis Presented to Addis Ababa University, Ethiopia 57p.
- United States Environmental Protection Agency (USEPA) (2005).Prevention, Pesticides and Toxic Substances: Mancozeb facts. Office of Pesticide Programs, US EPA, Washington, DC.
- William MA, Woodbury GW (1968). Specific gravity dry matter relationship and reducing sugar changes affected by potato variety, production area and storage. American Potato Journal 45(4):119-131

- Woldegiorgis G, Gebre E, Lemaga B (2008). Potato breeding. In: Woldegiorgis G., Gebre E., Lemaga B. (eds.). Root and Tuber Crops, the Untapped Resources. EIR, Addis Ababa, Ethiopia pp. 15-32.
- Yong-fu L, An-cheng L, Hassan MJ, Xinghua W (2006). Effect of phosphorus deficiency on leaf photosynthesis and carbohydrates partitioning in two rice genotypes with contrasting low P susceptibility. Rice Science 13:283-290.
- Zameer KM, Akhtar ME, Safdar MN, Mahmood MM, Ahmad S, Ahmed N (2010). Effect of source and level of potash on yield and quality of potato tubers. Pakistan Botany Journal 42(5):3137-3145.
- Zelalem A, Tekalign T, Nigussie D (2009). Response of potato (*Solanum tuberosum* L.) to different rates of N and P fertilization on Vertisol at Debre Berhan, in the central highlands of Ethiopia. African Plant Science Journal 3(2):16-24.

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Full Length Research Paper

## Short-term amelioration of soil properties and maize yield enhancement using animal wastes in degraded hydromorphic soils of Southeastern Nigeria

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Amending soil with animal wastes can be useful in food production as well as a means of waste disposal. It has been found to enhance physico-chemical properties and enhance yield. Poultry manure (PM), swine waste (SW), cow dung (CD), and sewage sludge (SS) were added to a hydromorphic ultisol (sandy loam, typic haplusult at the rate of 10 tha-1 (12 kgplot-1)). Maize hybrid (Oba super II) was used as test crop. Randomized Complete Block Design (RCBD) with four replications was used in laying the experiment. Data collected were analyzed using analysis of variance (ANOVA) and means were separated using Fishers' Least Significant Difference (F-LSD). Physical properties of soil influenced by animal wastes include bulk density, total porosity, hydraulic conductivity, gravimetric moisture contents, aggregate stability, and rheological characteristics. Amendments also enhanced soil organic matter, total nitrogen, available phosphorus, potassium, calcium, magnesium and sodium, pH, exchangeable acidity, cation exchange capacity, and base saturation were all higher relative to the unamended plots. Productivity studies revealed that maize growth was significantly affected by amendments especially PM. This was observed in height, leaf area index and yield higher relative to the control. Generally, the increase followed the order PM>SW>SS>CD. Animal wastes especially from poultry sources are recommended for soil amelioration and for increased crop yield in the area.

Key words: Cow dung, sewage sludge, swine waste, poultry manure, waste disposal.

#### INTRODUCTION

Sustainable agriculture is fast becoming the focus of the world today. Sustainable agriculture is a method of farming that is not only humane, environmentally friendly

and socially ethical, but can sustain itself. In the broadest sense, sustainable agriculture puts back into the earth what it takes out, making a cycle requiring no inputs from

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outside. For example, applying fertilizer derived from petroleum products is not sustainable, because the fertilizer was not produced within the agricultural cycle. Composted manure from animals on the farm would be an example of a sustainable fertilizer. Animal wastes may be suitable for amelioration of soils in the humid tropics which are characterized by poor native organic matter and low available nutrients productivity decline overtime when subjected to continous cultivation (Zingore et al., 2003).

In Nigeria, the use of mineral ferilizers is common among farmers, probably for its ability to enhance yield, low cost of application and transportation. However, mineral fertilizers overtime have proved to be scarce, cost intensive, capable of causing pollution and toxicity and incapable of achieving soil conservation needs (Kushwaha and Ochi, 1999). For example, Isherwood (2000) observed initial increase in yield with mineral fertilizer for only few years and followed by decrease in base saturation and acidification. Futhermore, Adeove et al. (2008) associated decreases in yield, increases in soil acidity, and nutrient imbalance to continous use of mineral ferilizers in tropical soils. Yield declines, pollution vulnerabilities and high cost of mineral fertilizers utilization has reawakened interests in organic wastes especially among the poor smallholder farmers.

The use of organic inputs such as crop residues and manures have great potential for improving soil productivity and crop yield through improvement of the soil physical, chemical, microbiological and nutrient supply (Abbasi et al., 2009). Ofori and Santana (1990) noted that cow dung improved the productivity of soil more than inorganic fertilizer owing to its slow release of nutrients. Organic manures can also increase water infiltration, water holding capacity, water content and aeration (McCauley et al., 2017). Many of the nutrients used by plants are held in organic manure until soil organisms decompose the material and release plant available nutrients. It is evident that a regular addition of organic manure is important as food for not only crops but also micro-organisms, insects, worms and other organisms. Erosion will be reduced and root penetration and tillage operation will be enhanced when the soil is well aggregated (USDA, 2003). Addition of soil organic matter is also an important soil conservation measure that accomplishes soil carbon sequestration and mitigation of climate change (McCauley et al., 2017).

Even though there are many studies dealing with organic manure application in the Abakaliki agro-ecology (Mbah et al., 2004; Mbah and Mbagwu, 2006; Nwite and Alu, 2017), very little is known about their effects on hydromorphic soils and yet such soils are common in the area. The objective of this study was to evaluate the suitability of animal wastes (poultry manure, swine wastes, cow dung, and sewage sludge) on soil physicochemical properties and yield components of maize (*Zea mays* L) in a degraded hydromorphic soil in Abakaliki,

Southeastern Nigeria.

#### **MATERIALS AND METHODS**

#### Description of study area

The study was carried out at the Teaching and Research Farm of the Faculty of Agriculture and Natural Resources Management, Ebonyi State University, Abakaliki. The area lies within latitude 06° 4¹N and longitude 08° 65¹E in the derived savanna zone of Southeastern Nigeria. Rainfall is bimodal; the rainy (April-October) and the dry season (November-March). There is usually a short break in August usually known as "August break". The total mean annual rainfall ranges from 1700 mm for minimum to 2000 mm for the maximum, respectively. The annual temperature is between 27 and 31°C, while relative humidity is between 60 and 80% during rainy season (Ofomata, 1975). The soil is hydromorphic and belongs to the order ultisol within the Ezzamgbo soil association derived from shale and classified as Typic Haplusult (FDALR, 1986).

#### Field work

The study was conducted in a land area of approximately 0.017 ha  $(13 \times 13 \text{ m}^2)$ . The field was cleared manually of existing vegetation and debris removed. The area was demarcated into plots that measured  $2 \times 2 \text{ m}^2$  with 0.5 m space. The Randomized Complete Block Design (RCBD) was used in the experimental design. Animal wastes consisting of poultry manure (PM), swine waste (SW) and cow dung (CD) obtained from the Animal Science section of Ebonyi State University Abakaliki and sewage sludge (SS) were obtained from sewage treatment plant of University of Nigeria, Nsukka. The animal wastes were dried, crushed, analyzed for their respective nutrient components and incorporated into the soil at 20 cm depth prior to planting. The treatments were 10 tha  $^{-1}$  (4 kg/plot) of each animal wastes (PM, SW, CD and SS) and a control. They were replicated four times to give a total of twenty experimental plots.

Maize (*Zea mays* L. var Oba super II) was planted as test crop two weeks after incorporation of treatments. The seed rate was two seed per hole at a spacing of  $25 \times 75$  cm<sup>2</sup> and depth of 5 cm. The seedlings were thinned down to one per hole at two weeks of germination and weeding was bi-weekly.

#### Agronomic data

Twelve plants constituting 25% of plant population per plot were tagged and used for agronomic measurements. Plant height was measured with metric ruler from tallest leaf of a plant to base every two weeks till tasseling. The grain yield was determined by harvesting the cobs after drying of husks. The husks were removed, cobs shelled and maize grains were further dried and grain yield was adjusted to 14% moisture content.

#### Laboratory studies

Selected soil physical and chemical properties were determined after soil samples were collected with core samplers and augers, respectively auger at 0 to 20 cm. Auger samples were composited, dried, ground, seived with 2 mm-mesh seive and labelled at pre- and post-planting. Bulk density was determined using the method described by Gee and Or (2002). Total porosity determination was done as described by Obi (2000). The method of Stolte (1997) was used to determine saturated hydraulic conductivity (Ks). Gravimetric

**Table 1.** Chemical composition of amendments used for the study.

Manure	pH (KCI)	O.C (%)	TN (%)	Av.P (mgkg <sup>-1</sup> )	Exch. K (cmol kg <sup>-1</sup> )	Exch. Ca (cmol kg <sup>-1</sup> )	Exch. Na (cmol kg <sup>-1</sup> )	Exch Mg (cmol kg <sup>-1</sup> )
PM	7.5	23.0	3.67	0.4	0.55	3.27	0.33	1.52
SW	7.0	20.2	2.45	0.35	0.63	3.19	0.23	1.58
CD	7.0	26.2	3.21	0.34	0.47	2.81	0.11	1.22
SS	5.91	26.2	2.76	0.16	0.48	2.91	0.28	1.39

PM: Poultry manure; SW: swine wastes; CD: cow dung; SS: sewage sludge; O.C: organic carbon; TN: total nitrogen; Av. P: available phosphorus.

moisture content determination was carried out as described by Obi (2000). Particle size distribution (fraction of sand, silt, and clay) was determined using hydrometer method (Gee and Or, 2002) with NaOH as dispersant. Mean weight diameter was determined using the method described by Obi (2000). Sower (1965) penetrometer and classical techniques were used to determine liquid limit and plastic limit, respectively. Soil pH was determined in 1M KCl of 1:2.5 soil/water ratio. Organic carbon was determined by the Walkley and Black dichromate oxidation method (Nelson and Sommers, 1982). Organic matter was estimated as organic carbon x 1.729 (Odu et al., 1986). And total nitrogen (TN) by Kjeldahl method (Bremner and Mulvaney, 1982). Available phosphorus by Bray and Kurtz (1945) (Bray II) method. After extraction with 1N ammonuim acetate solution, exchangeable Na and K were determined by the Flame photometry and Ca and Mg by an EDTA titration method. Exchangeable acidity ( $Al^{3+}$  +  $H^+$ ) was determined by the KCl displacement method and cation exchange capacity (CEC) was obtained by the ammonuim acetate technique. Percentage base saturation was calculated as total exchangeable bases divided by effective CEC and multiplied by 100. All laboratory analysis was conducted at the Soil Science Research Laboratory, University of Nigeria Nsukka.

#### Data analysis

All data collected were analyzed statistically using the analysis of variance technique (ANOVA) and Fishers least significant difference (Steel and Torrie, 1980) was used to separate means and significance was accepted at 5% probability level.

#### **RESULTS AND DISCUSSION**

## Chemical composition of amendments used for the study

Table 1 shows the chemical composition of the animal wastes used for soil amendment. Total N was higher (3.68%) in poultry manure following the order PM>CD>SS>SW. Sewage sludge was slightly acidic (5.91) while other wastes were neutral to alkaline. Although values of available P were low in the animal wastes based on Landon (1991), it was highest in poultry manure (0.4 mgkg<sup>-1</sup>). The exchangeable Ca values were 3.72, 3.19, 2.81 and 2.91 cmolkg<sup>-1</sup> for poultry manure, swine waste, cow dung and sewage sludge, respectively and higher than Mg, K and Na values in the wastes.

Exchangeable Mg and K were dominant in swine waste (1.58 and 0.63 cmolkg<sup>-1</sup>), respectively as compared to other wastes.

#### Soil properties prior to amendment

The properties of the soil at the initiation of the study are shown in Table 2. Sand fraction was dominant compared to clay and silt fractions leading to sandy clay loam textural class. The pH was slightly acidic (5.2) and organic carbon content (1.8%) was also low based on rating by Landon (1991). Total nitrogen (0.11%), available phosphorus (26 mgkg<sup>-1</sup>), cation exchange capacity (8.6 cmolkg<sup>-1</sup>) were rated low (Landon, 1991). Exchangeable calcium and magnesium dominated the exchange sites.

#### Effect of treatments on soil physical properties

Table 3 shows the physical properties as influenced by animal wastes. In all treatments, the texture was sandy clay loam which may be attributed to nature of parent materials and high rainfall that could favor washing away and leaching of silt-sized and clay-sized fractions (Igwe et al., 1999; Akamigbo, 2010). The application of animal wastes significantly (P<0.05) decreased bulk density and increased total porosity. The implication of a lowered bulk density and increased total porosity are ease in root penetration, downward movement of water, more soil retention, availability for greater water use water efficiency by crops and lowered risks of compaction (Ogbodo and Chukwu, 2012; Nwite and Okolo, 2016). Bulk density was lower in SS (1.45 gcm<sup>-3</sup>) amended plots relative to CD (1.51 gcm<sup>-3</sup>), PM (1.54 gcm<sup>-3</sup>) and SW (1.62 gcm<sup>-3</sup>) plots. This reflects the role that the different organic waste decomposition plays in soil loosening. However, saturated hydraulic conductivity obtained in PM amended plots were higher relative to other amendments, following the order PM>CD>SS>SW>C. PM relative to the control increased water transmission through the soil by 61.9%. Similarly, gravimetric moisture contents of PM plots were higher relative to the amended and

**Table 2.** Pretreatment soil (0-20 cm) properties.

Particle size distribution	Values
Clay%	21
Silt%	20
Sand%	59
Textural Class	SCL
Soil pH (KCI)	5.2
Exch. Ca (cmol kg <sup>-1</sup> )	4.1
Exch. Mg (cmol kg <sup>-1</sup> )	2.8
Exch. Na (cmol kg <sup>-1</sup> )	0.03
Exch. K (cmol kg <sup>-1</sup> )	0.1
CEC (cmol kg <sup>-1</sup> )	8.6
Exch. Acidity (cmol kg <sup>-1</sup> )	1.36
TN (%)	0.11
OC (%)	1.8
Av.P (mgkg <sup>-1</sup> )	26

O.C: organic carbon; TN: total nitrogen; Av. P: available phosphorus; Exch Ca: exchangeable calcium; Mg: magnesium; Na: sodium; K: potassium.

**Table 3.** Physical and properties of the soil as influenced by wastes.

Treatment	%Sand	%Silt	%Clay	Tex	BDg (cm <sup>-3</sup> )	TP (%)	HC (cmh <sup>-1</sup> )	GMC (%)	AS (%)	SA (%)	MWD (%)	LL	PL
Control	59	20	21	SCL	1.66	36.3	16	13.3	10.8	2.3	1.6	19.8	15.2
PM	59	23	18	SCL	1.54	42.5	41	19.0	14.7	4.7	2.5	22.5	17.7
SW	58	23	19	SCL	1.62	37.8	28	15.6	11.2	3.6	2.4	21.2	16.3
CD	58	22	20	SCL	1.51	41.8	36	14.5	13.3	3.6	2.4	20.9	18.1
SS	60	21	19	SCL	1.45	41.3	30	16.8	11.1	3.4	2.4	21.5	17.0
FLSD (0.05)	NS	NS	NS	-	0.07	3.3	NS	NS	NS	NS	NS	22.2	17.9

Tex: Soil texture; SCL: sandy clay loam; BD: bulk density; TP: total porosity; HC: hydraulic conductivity; GMC: gravimetric moisture content; AS: aggregate stability; SA: state of aggregate; MWD: mean weight diameter; LL: liquid limit; PL: plastic limit.

unamended plots. There was no significant difference between the control and amended plots in terms of aggregate stability and state of aggregation. Notwithstanding, what seems obvious are slight increase with amendment following the order PM>CD>SW>SS>C. This corroborates the findings of Wang et al. (2016) who observed that organic matter is an indispensible component in soil aggregation. Similarly, there were no significant differences among the treatments in terms of mean weight diameter. However, PM recorded the highest values (2.5%) as compared to other amendments and control (1.6%). This indicates that animal wastes enhanced mean weight diameter of soil.

The liquid and plastic limits of amended soil were significantly (P<0.05) increased following amendments. Higher plasticity values were recorded in CD (18.1), PM (17.7), SS (17.0) and SW (16.3) amended plots as compared to the control (15.2). The implication of this is that moisture contents of the soils were improved

following the amendments.

#### Effect treatments on soil chemical properties

## Soil pH, organic matter, total nitrogen and available phosphorus

Soil pH was not significantly (P>0.05) affected following organic waste amendment as shown in (Table 4). The highest value of 6.1 was obtained in plots amended with poultry manure while the lowest was 5.2 from the control. The increase in pH due to animal waste amendment have been widely reported in literature (Darmordy et al., 1983; Nwite et al., 2016).

Amendments increased P significantly (P<0.05). The highest value was obtained on poultry manure amended plots (40 mgkg<sup>-1</sup>) as compared to the lowest value (28 mgkg<sup>-1</sup>) recorded in the control (Table 4). This agrees

**Table 4.** Chemical properties of soil at post Harvest.

Treatment	рН	Av. P	TN	ОМ	C:N	Ca	Mg	K	Na	EA	CEC	DC (0/)
Treatment	(KCI)	(mgkg <sup>-1</sup> )	(%)	(%)	ratio			Cn	nolkg <sup>-1</sup>			BS (%)
С	5.2	28	0.11	2.0	10	4.1	2.8	0.1	0.03	1.36	8.4	80
PM	6.1	40	0.13	2.3	11	6.1	3.2	0.20	0.07	1.22	10.9	86
SW	5.8	28	0.09	2.0	13	5.1	2.8	0.13	0.06	1.30	10.2	83
CD	5.9	30	0.12	2.2	11	4.9	2.8	0.11	0.06	1.28	9.4	83
SS	5.7	36	0.11	2.1	11	4.6	2.8	0.13	0.08	1.34	9.5	81
FLSD (0.05)	NS	7.5	NS	NS	NS	NS	NS	NS	0.02	NS	1.65	NS

C: Control; PM: poultry manure; SW: swine wastes; CD: cow dung; SS: sewage sludge.

**Table 5.** Effect of amendments on growth and yield of maize.

Treatment	Plant height (cm)	Leaf area index	Grain yield (tha <sup>-1</sup> )
С	113.2	344	0.5
PM	161.7	519	1.6
SW	137.5	463	1.4
CD	129.4	397	0.9
SS	136.1	387	1.2
FLSD (0.05)	20.3	154	0.2

C: Control; PM: poultry manure; SW: swine wastes; CD: cow dung; SS: sewage sludge.

with earlier findings by Adeleye and Ayeni (2009) that P content is increased by application of animal wastes.

Total N exhibited non-significant effect following wastes amendments. Like P, the highest value of 0.13% was obtained in poultry manure amended plots, while the lowest value (0.11%) was recorded in the control. There was no significant difference (P>0.05) among the treatments. The increase in total N of amended plots is similar to the effects observed by Khaliq and Abbasi (2015) and attributed to build up of organic matter in the soil.

There was no significant effect of amendments on soil organic matter. However, plots amended with poultry manure gave the highest values of 2.3%, while CD, SS and SW amended plots gave 2.2%, 2.1% and 2.0 as compared to the control plots 1.9%. This is consistent with reports of Wang et al. (2014) soil organic matter was higher in organic waste amended soils and is dependent on its nature and rate of its decomposition by the microbial community. PM contains both solid and liquid excreta and tends to mineralize faster as compared to wastes from other sources where urine is lost-resulting in high solid excreta (Amanullah et al., 2010). This explains the low C:N ratio of PM amended soils as an indicative faster release of nitrogen.

#### Exchangeable bases and exchangeable acidity

Amendments led to an increase in soil CEC especially in

PM plots (Table 4). This implies better supply of nutrients, hence improved fertility in amended plots. The higher values in poultry manure amended plots for Ca<sup>2+</sup>, K<sup>+</sup> and Mg<sup>2+</sup> has been observed by other reseachers (Hue and Lucidine, 1999; Adeniyi and Ojeniyi, 2005). There is probably an increase in the amount of Ca2+ derived from CaCO<sub>3</sub> due to formation of organic acids. Increased Ca<sup>2+</sup> will result in improved soil structure by forming cationic bridges between clay and soil organic matter (David and Dimitrios, 2002). Mg<sup>2+</sup> content tended to remain unaltered in all plots excluding the PM amended. The effect of amendments on Na content was significantly increased. SS amended plots had the highest values for Na<sup>+</sup> with value of 0.08 cmolkg<sup>-1</sup>, while the control plots had the lowest values. This corroborates early findings (Basta, 1996), that all wastes have high soluble salts in variable quantities and excessive salts in soils (natural or applied) can have detrimental effects on plants growth.

Exchangeable acidities were not significant (P>0.05). Sewage sludge had the highest value of 1.86 cmolkg<sup>-1</sup>. The changes may be due to Al<sup>3+</sup> from soil exchange site acted upon by decomposing waste-humus having a weak acid nature (Adediran et al., 2003).

#### Effect of treatments on growth and yield components

Plant height was significantly (P<0.05) increased (Table 5) with the application of the amendments. The highest

result of 161.73 cm was 43% higher than the control plots. This was also observed in the leaf area index (51%) and yield (220%) higher relative to the control. It followed the order PM>SW>SS>CD for plant height, leaf area index and yield. This agrees with Mbah (2008) and Nwite et al. (2016) who observed that organic manure improves crops yield and is source-dependent.

#### Conclusion

The study shows that animal wastes enhanced soil properties and maize yield of hydromorphic ultisols in Abakaliki agro-ecology. The enhancement of soil properties and yield followed the order PM>SW>SS>CD. Poultry manure having low C:N ratio had the ability to release faster, hence, performing better than other animal wastes in the short-term. Cow dung, swine wastes and sewage sludge may have greater residual benefits. The application of sewage sludge may be beneficial in certain soil deficiency, but however has toxicity potentials. The application of these wastes at the rate used in this study will lead to restoration of degraded soils, thus ensuring that soil is healthy enough to perform its functions for agricultural sustainability. More studies are however needed using variable quantities and combination of wastes in hydromorphic soils. A long-term study of the wastes in order to bring out a cumulative effect of the amendments will also be a very useful practice.

#### **CONFLICT OF INTERESTS**

The authors have not declared any conflict of interests.

#### **REFERENCES**

- Abbasi MK, Mushtaq A, Tahir MM (2009). Cumulative effects of white clover residues on the changes in soil properties, nutrient uptake, growth and yield of maize crop in the sub-humid hilly region of Azad Jamnu and Kashmir Pakistan. African Journal of Biotechnology 8(22):6191.
- Adediran JA, Taiwo LB, Sobulo RA (2003). Effect of organic wastes and method of compositing on compost, nutrient maturity, composition of compost and yield of two vegetable crops. Journal of Sustainable Agricultural 22:95-109.
- Adeleye EO, Ayeni LS (2009). Effects of soil preparation methods and organic wastes on soil nutrient status and yield of maize (*Zea mays*) on an alfisol of southwest Nigeria, American-Eurasian Journal of Sustainable Agriculture 3(3):460-467.
- Adeniyi ON, Ojeniyi SO (2005). Effect of poultry manure, NPK 15-15-15 and combination of their reduced levels on maize growth and soil chemical properties. Nigerian Journal of Soil Science 15:34-41.
- Adeoye GO, Adeoluwa OO, Ojelade M, Sridhar E, Makinde EA, Olowoake AA (2008). Comparative evaluation of organo-mineral fertilizers and mineral fertilizers (NPK) on yield quality of maize (Zea mays (L) Moench). Nigerian Journal of Soil Science 18:132-137.
- Akamigbo FOR (2010). Soils: Fundamental Methods of Soil Resource Survey, Classification, Interpretation, Interpretation and Application. Published and printed by University Press Ltd. Bookshop/Bank building complex, UNN.
- Amanullah MM, Sekar S, Muthukrisnan P (2010). Prospects and

- potential of poulty manure. Asian Journal of Plant Science 9:172-192.
- Basta NT (1996). Soil amendments: Impacts in biotic systems. Soil Science 161(12):885-886.
- Bray RH, Kurt LT (1945). Determination of total, organic and available forms of phosphorus in soils. Soil Science 59:39-45.
- Bremner JM, Mulvaney CS (1982). Nitrogen total. In Page, A.L.(ed) Methods of soil analysis. No 9. Part 2: Chemical and Microbiological properties, 2nd edn. Am Soc Agron Inc, Madison, WI, USA.
- Darmordy RG, Foss JE, Mc itosh M, Wolf DC (1983). Municipal sewage sludge compost amended soils: some spatiotemporal treatment effects. Journal of Environmental Quality 12:2.
- David R, Dimitrios P (2002). Diffusion and cation exchange during the reclamation of saline-structured soils. Geoderma 107:271-279.
- FDALR (1986) Reconnaissance soils survey of Anambra State. Nigeria Soils report FDALR Kaduna.
- Gee GW, Or D (2002). Particle size analysis. In: Dane, J. H. and Topp, G. C. (eds). Methods of soil Analysis. Part 4; physical method. Soil Science Society American 5:255-293.
- Hue NV, Licudine DL (1999). Amelioration of subsoil acidity through surface applications of organic manures. Journal. Environmental Quality 28:623-632.
- Igwe CA, Akamigbo FOR, Mbagwu JSC (1999). Chemical and mineralogical properties of soils in Southern Nigeria in relation to aggregate stability. Geoderma 92:111-123.
- Isherwood KF (2000). Fertilizer use and the Environment, International Fertilizer Industry Association Revised Edition. Paris, France.
- Khaliq A, Abbasi MK (2015). Improvement in the physical and chemical characteristics of degraded soils supplemented with organic-inorganic amendments in the Himalayan region of Kashmir, Pakistan. Catena 126:209-219.
- Kushwaha S, Ochi JE (1991). The economic and social implication of using animal excreta as fertilizer and feedstuff: profitability and environmental protection model. Journal on sustainable Agriculture 14(1):81.
- Landon JR (1991). Booker, Tropical soil Manual: A handbook for soil survey and agricultural land evaluation in tropics and subtropics. New York, USA. John Wiley and Sons, Inc. Third Avenue.
- Mbah CN (2008). Contribution of Organic amendments to exchangeable Potassium percent and soil nitrate concentration in an Ultisol and their effect on maize (*Zea mays L*) grain yield. Agroscience Journal of Tropical Agricultural, Food, Environment and Extention 7(3):206-210.
- Mbah CN, Mbagwu JSC (2006). Effect of animal waste on physiochemical properties of a Dystric Leptosol and maize yield in South Eastern Nigeria. Nigerial Journal of soil Science 16(1):96-103.
- Mbah CN, Mbagwu JSC, Onyia VN, Anikwe MAN (2004). Effect of application of biofertilizer on soil densification, total porosity, aggregate stability and maize grain yield in dystric leptosol at Abakaliki. Nigeria Journal of Science Tecnology 10:74-85.
- McCauley A, Jones C, Olson-Rutz K (2017). Soil pH and organic matter. Nutrient Management Module No. 8, 4449-8. Montana State University/ Extension.
- Nelson DW, Sommers LW (1982). Total organic carbon and organic matter. In: Pg (eds) Methods of soil analysis, part 2.Chemical and microbiological properties. Agronomy monograph, No. 7 Madison W, USA.
- Nwite JN, Alu MO (2017). Potentials of animal wastes in productivity of a sandy loam in Abakaliki, Southeastern Nigeria. American-Eurasian Journal of Sustainable Agriculture 11(4):1-7.
- Nwite JN, Aya FC, Okolo CC (2016). Remediation of Crankcase Oil Contaminated Soil with Different Sources of Manure in Abakaliki, South East, Nigeria. Journal of Bioremediation Biodegration 7:350. DOI:10.4172/2155-6199.1000350.
- Nwite JN, Okolo CC (2016) Soil water relations of an Ultisol amended with agro-wastes and its effect on grain yield of maize (*Zea Mays* L.) in Abakaliki, Southeastern Nigeria. European Journal of Scientific Research 141(2):126-140.
- Obi ME (2000). Soil Physics. A compendium of lectures. Nsukka, Atlantic Publishers.
- Odu CT, Babalola IO, Udo EJ, Ogunkunle AO, Bakare TA, Adeoye GO (1986). Laboratory manual for agronomic studies in soil plant and microbiology. Department of Agronomy, University of Ibadan, Nigeria

- P 83.
- Ofori CS, Santana R (1990). Fertilizer their potential and use in Africa agriculture in organic matter management and tillage in sub-humid Africa- IIBSRAM PROC 10:213-219.
- Ofomata GEK (1975) Nigeria in maps, Eastern states. In G.E.K. Ofomata E.d Ethiopia pub house, Benin City Pp. 45-46.
- Ogbodo EN, Chukwu GO (2012). Soil fertility evaluation of selected Aquic Haplustalfs in Ebonyi State, Southeast Nigeria. Nigeria Journal. of Soil Science 22(1)97-102.
- Sower GF (1965) Consistency method of soil analysis, part 1 1<sup>st</sup> (Ed) in: C.A Black *et al* American Society of Agronomy, Madison, Ulis Cousin pp. 391-399.
- Steel RGD, Torrie JH (1980). Principles and procedures of statistics. A biometric approach. 2nd ed. McGraw Hill, Book Co. Inc. New York pp. 1-3.
- Stolte J (1997). Manual of soil physical measurement. Version 3 Wageningen, DLO Winand Staring Centre for Integrated Land. Soil and Water Research, Technical Document P 37.
- United State Development Agency (USDA) (2003). Organic matter management and soil conservation into agricultural service information bulletin No 229.

- Wang L, Sun X, Li S, Zhang T, Zhang W, Zhai P (2014). Application of organic amendments to a coastal saline soil in North China: effects on soil physical and chemical properties and tree growth. PLoS ONE 9(2):e89185. doi:10.1371/journal.pone.0089185.
- Wang J, Yang W, Bi Y, Li Z, Ma R (2016). Estimating the influence of related soil properties on macro- and micro-aggregate stability in ultisols of south-central China. Catena 137:545-553.
- Zingore S, Mafongoya P, Myamagafota P, Giller KF (2003). Nitrogen mineralization and maize yield following application of tree prunning to a sandy soil in Zimbabwe. Agroforestry system 57:119-211.

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